



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Genetic mapping of the female mimic morph locus in the ruff

Citation for published version:

Farrell, LL, Burke, T, Slate, J, McRae, SB & Lank, DB 2013, 'Genetic mapping of the female mimic morph locus in the ruff', *BMC Genetics*, vol. 14, pp. 109. <https://doi.org/10.1186/1471-2156-14-109>

Digital Object Identifier (DOI):

[10.1186/1471-2156-14-109](https://doi.org/10.1186/1471-2156-14-109)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

BMC Genetics

Publisher Rights Statement:

Copyright © 2013 Farrell et al.; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



RESEARCH ARTICLE

Open Access

Genetic mapping of the female mimic morph locus in the ruff

Lindsay L Farrell^{1,2*}, Terry Burke¹, Jon Slate¹, Susan B McRae³ and David B Lank²

Abstract

Background: Ruffs (*Aves: Philomachus pugnax*) possess a genetic polymorphism for male mating behaviour resulting in three permanent alternative male reproductive morphs: (i) territorial 'Independents', (ii) non-territorial 'Satellites', and (iii) female-mimicking 'Faeders'. Development into independent or satellite morphs has previously been shown to be due to a single-locus, two-allele autosomal Mendelian mode of inheritance at the *Satellite* locus. Here, we use linkage analysis to map the chromosomal location of the *Faeder* locus, which controls development into the Faeder morph, and draw further conclusions about candidate genes, assuming shared synteny with other birds.

Results: Segregation data on the *Faeder* locus were obtained from captive-bred pedigrees comprising 64 multi-generation families ($N = 381$). There was no evidence that the *Faeder* locus was linked to the *Satellite* locus, but it was linked with microsatellite marker *Ppu020*. Comparative mapping of ruff microsatellite markers against the chicken (*Gallus gallus*) and zebra finch (*Taeniopygia guttata*) genomes places the *Ppu020* and *Faeder* loci on a region of chromosome 11 that includes the *Melanocortin-1 receptor (MC1R)* gene, which regulates colour polymorphisms in numerous birds and other vertebrates. Melanin-based colouration varies with life-history strategies in ruffs and other species, thus the *MC1R* gene is a strong candidate to play a role in alternative male morph determination.

Conclusion: Two unlinked loci appear to control behavioural development in ruffs. The *Faeder* locus is linked to *Ppu020*, which, assuming synteny, is located on avian chromosome 11. *MC1R* is a candidate gene involved in alternative male morph determination in ruffs.

Background

Evolving and maintaining genetic polymorphisms responsible for large phenotypic differences remains a subject of interest, despite >70 years of study (e.g. [1,2]). Genomic methods now enable polymorphisms to be described down to the genetic and molecular expression levels (e.g. [3]). Ruffs (*Philomachus pugnax*) possess three distinct permanent alternative male reproductive morphs that differ in territorial lekking behaviour, plumage colour, and size: dark-plumed territorial 'Independents', white-plumed non-territorial 'Satellites' and small female mimic 'Faeders' that lack display plumage and behaviour [4-7]. Development into independent or satellite morphs has been previously shown to be due to a genetic polymorphism consistent with a single-locus, two-

allele autosomal Mendelian mode of inheritance at the *Satellite* locus, with a dominant *S* allele producing satellites [8,9]. Genetically, independent males are homozygous recessive at the *Satellite* locus and *ca* 90% of satellites should be heterozygotes [8]. Remarkably, only as recently as 2006, a third morph was discovered: faeder males resemble large females, completely lacking any ornamental breeding plumage during the breeding season [5] (Figure 1). It was recently reported that a dominant allele controls development into both faeders and diminutive females, coined 'faeder females' [10]. Whether the faeder allele is at the same *Satellite* locus, or a separate locus, has yet to be determined, as more detailed pedigree-based genetics of the newly discovered morph are not yet available.

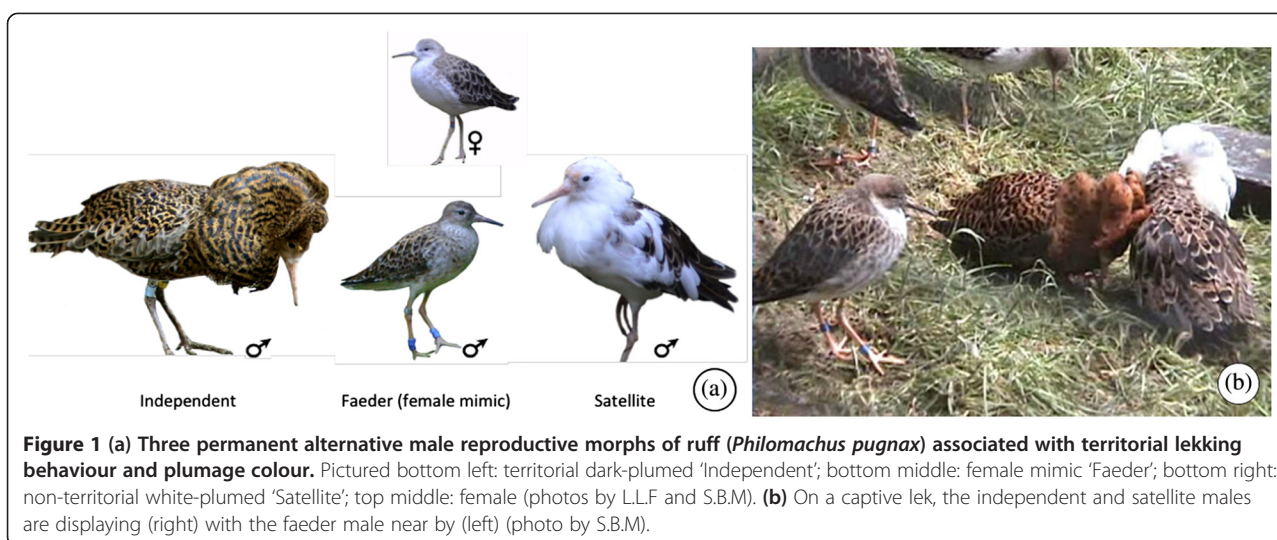
Prior to each breeding season, independent and satellite males grow ornamental plumage that includes a feather 'ruff' and 'head tufts', which are each individually distinctive in colour and pattern and fixed for life [11,12]. At leks, independents establish and defend small breeding courts where they perform a variety of territorial threat displays

* Correspondence: lfarrell@sfu.ca

¹Department of Animal and Plant Sciences, University of Sheffield, Sheffield, S10 2TN, UK

²Department of Biological Sciences, Simon Fraser University, Burnaby, BC V5A 1S6, Canada

Full list of author information is available at the end of the article



and fight against other independents. The white-plumaged satellites do not hold territories, are rarely aggressive, and are actively courted into co-displaying on courts held by independents, apparently due to female preference for male-male cooperation on leks [4,6,7,13,14] and a high rate of polyandry [15]. In contrast to both classes of ornamented males, faeder males grow breeding plumage that is similar to that of females—lacking display feathers—and aggregate close to displaying males to ‘sneak’ copulations with females and interfere with copulation attempts by other males ([5]; Lank *et al.* unpublished) (Figure 1). Females believed to be carrying the dominant *Faeder* allele form a distinct small size mode [10]. Normal-sized females carrying the dominant *Satellite* allele can be identified from the phenotype ratios of their male offspring when mated to independent males, and/or confirmed with observations of behaviour and ornamental plumage growth when implanted with testosterone [9].

Recently, a microsatellite linkage map for the ruff was constructed, identifying seven linkage groups and a further five single-marker loci homologous to locations on known chicken (*Gallus gallus*) and zebra finch (*Taeniopygia guttata*) chromosomes [16]. As a step towards identifying the genes underlying the morph polymorphisms, we attempted to map the causal satellite and faeder loci by using linkage analysis to identify markers that co-segregated with each morph type in a pedigreed and phenotyped breeding population.

Methods

Pedigree, phenotype, and microsatellite information were available from 381 individuals from a captive population of ruffs spanning fourteen breeding years and comprising 64 families ($N = 381$ individuals, [10,16]). In total, 167 individuals were included for the *Satellite* locus: 129

assigned as independents (120 males, 9 females), 38 satellites (35 males, 3 females) and 381 individuals for the *Faeder* locus: 43 faeders (24 males, 19 females) and 338 non-faeders (155 males, 183 females).

This research was conducted at Simon Fraser University under approval of the Animal Care Committee.

Linkage analysis

Separate autosomal genetic models for the two male behavioural polymorphisms (*Satellite versus Independent*; *Faeder versus Not Faeder*) were tested in CRIMAP v.2.4 [17] using phenotypic and pedigree data to assign putative genotypes separately for both the *Satellite* and *Faeder* loci. For the *Satellite* locus: independent males ($N = 120$) were coded as homozygous recessive (ss) and satellite males ($N = 35$) coded with the dominant S allele (S_{-}), with faeders not coded at this locus. A small number of females ($N = 12$) were assigned a satellite or independent behavioural morph and putative genotype based on pedigree analysis of their male offspring morph ratios when mated with an independent male (Lank *et al.* unpublished). Females mated with an independent male that produced mixed offspring were designated as heterozygotes (Ss , $N = 3$), and females with a high number of offspring ($N = 11-22$) who failed to produce any satellites when mated with independents were designated as homozygous recessive at the *Satellite* locus (ss , $N = 9$). In the majority of cases, these morph assignments were confirmed with testosterone-induced behavioural data [9]. For the *Faeder* locus: both independent and satellite males were coded as homozygous recessive (ff , $N = 155$) and faeder males as (F_{-}), indicating that they carry at least one copy of the F allele ($N = 24$) [10]. Since the faeder frequency in natural populations is *ca* 1% [5,18-20], the probability of observing homozygous faeders in the wild is low. Faeders in the captive

population were derived from 2 wild-caught founders. Both of these males produced both faeder and non-faeder offspring when mated exclusively with females from non-faeder lineages, as did their sons. No faeder daughters are included as mothers in these analyses. For females, phenotypic assignments as 'faeder females' ($N=19$) were made through principal component analysis of size distributions based on tarsus, culmen, and minimum mass [10]. All non-faeder females ($N=183$) were coded as homozygous recessive (ff), and faeder females coded as (F_-) for similar reasons as were the males.

A test for linkage between the *Satellite* locus and *Faeder* locus, and all microsatellite markers ($N=58$) used in the ruff microsatellite linkage map [16], was performed by means of the two-point function in CRIMAP, with a LOD score >3.0 being taken as evidence of linkage. The *Satellite* and *Faeder* loci were first run separately, then together in CRIMAP. We used comparative mapping [21,22] of microsatellite markers used in the ruff microsatellite linkage map [16] against the chicken and zebra finch genome assemblies to search for possible candidate genes in the genomic location close to any microsatellites that were linked to the ruff *Faeder* locus.

Results and discussion

No linkage was detected between the *Satellite* and *Faeder* loci, and the *Satellite* locus was unlinked to any other marker in twopoint analysis. The latter result may be due in part to the low number of satellites with heterozygous genotypes and high number of independents contained within the pedigree, resulting in a small number of informative meioses at the target *Satellite* locus. Out of the total 167 individuals with inferred genotypes at the *Satellite* locus, 129 of these were independents and 38 were satellites. The non-linkage of the two behavioural loci, *Satellite* and *Faeder*, to the same marker or, more importantly, to each other, indicates that two independent loci determine alternative morph development in ruffs. Additional genotyping of satellite individuals and/or more detailed pedigree data will further test this two-locus model.

Several species with three heritable alternative mating phenotypes have been described (e.g., [23]), but explicit mendelian models have been best tested for the marine isopod *Paracerceis sculpta* [24], for which a 1-locus 3-allele model with hierarchical dominance was supported. Remarkably, alleles coding for 'alternative' morphs in these other systems are dominant to those of the presumed ancestral allele, as they are in the ruff [10]. In the ruff, this suggests a sequence for invasion by these derived morphs, with faeders following satellites.

In the twopoint analyses, the *Faeder* locus was strongly linked to microsatellite marker *Ppu020* with a LOD score 8.24 and recombination fraction of 0.03. This locus was not placed on the ruff linkage map but comparative

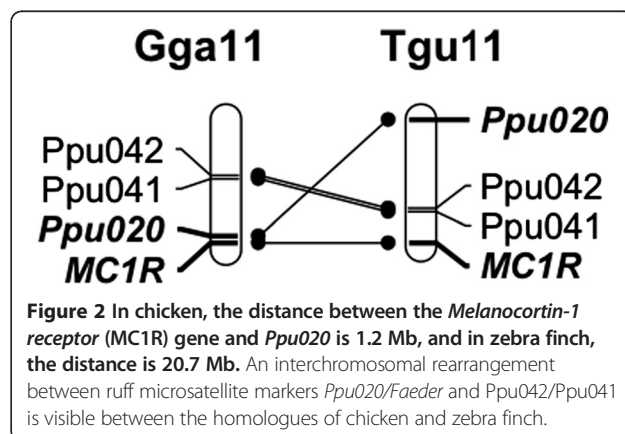
mapping has shown it to be on chromosome 11 [16]. Further linkage analysis with microsatellite markers on chromosome 11 was not possible, however, due to the small number of markers genotyped on this chromosome in the ruff linkage map [16].

By comparative mapping [21,22] of ruff microsatellite markers [25] to the chicken and zebra finch genome assemblies (Figure 2), an obvious candidate locus was identified. The *Melanocortin-1 receptor* (*MC1R*) gene, an important pigment-regulating gene in birds and numerous other vertebrates, is located on chromosome 11 in both species. In chicken, the distance between *Ppu020* and *MC1R* is 1.2 Mb and, in zebra finch, the distance is 20.7 Mb (Figure 2).

Although ruff microsatellite *Ppu020* is not included in the ruff linkage map for chromosome 11, two further ruff microsatellite loci have been assigned to this chromosome by *in silico* comparative mapping [25]. Comparison of the locations of these markers in the zebra finch and chicken genomes indicates that there was an intrachromosomal rearrangement of this region of chromosome 11 in an unknown lineage since the divergence of the ancestors of chicken and zebra finch (Figure 2). Therefore, inferring the physical distance between *MC1R* and the *Faeder* locus in ruffs is not straightforward, especially as no species in the ruff's avian superorder (the Charadriiformes) has yet been the subject of a full genome sequencing project.

Conclusion

Regardless of the precise location of *MC1R* in ruffs, we conclude that this gene and those in proximity to it are candidates for the *Faeder* locus. Melanin-based colouration has previously been shown to be associated with morphology, physiology, life-history strategies and behaviour in several bird species (e.g., [26-28]), including ruffs, as well as having correlated fitness-related effects in other vertebrates [29,30].



Competing interests

The authors declare no competing interests.

Authors' contributions

LLF designed the study, performed the laboratory work, completed the data analysis, and drafted and revised the manuscript. DBL, TB and JS helped with the design of the study, the interpretation of the data, and with drafting and revision of the manuscript. JS and TB conceived coding the faeder behaviour locus into the linkage map. SBM contributed to the development of the ruff pedigree and quantifying faeder phenotypes with DBL. All authors read and approved the final manuscript.

Acknowledgements

During this work, LLF was a PhD student in the department of Biological Sciences at Simon Fraser University. The laboratory work was performed at the University of Sheffield and supported by a UK Biotechnology and Biological Sciences Research Council grant to TB and JS. The captive ruff colony was supported by a Natural Sciences and Engineering Research Council of Canada grant (NSERC; to DBL), and LLF was supported by an NSERC PGS-D3.

Author details

¹Department of Animal and Plant Sciences, University of Sheffield, Sheffield, S10 2TN, UK. ²Department of Biological Sciences, Simon Fraser University, Burnaby, BC V5A 1S6, Canada. ³Department of Biology and Center for Biodiversity, East Carolina University, Greenville, NC 27858-4353, USA.

Received: 29 July 2013 Accepted: 11 November 2013

Published: 20 November 2013

References

- Chevin LM, Lande R: Evolution of discrete phenotypes from continuous norms of reaction. *Am Nat* 2013, **182**:13–27. doi:10.1086/670613.
- Dobzhansky T: *Genetics of the Evolutionary Process*. New York: Columbia University Press; 1970.
- Cheviron ZA, Whitehead A, Brumfield RT: Transcriptomic variation and plasticity in rufous-collared sparrows (*Zonotrichia capensis*) along an altitudinal gradient. *Mol Ecol* 2008, **17**:4556–4569.
- Hogan-Warburg AJ: Social behavior of the ruff (*Philomachus pugnax*). *Ardea* 1966, **54**:108–229.
- Jukema J, Piersma T: Permanent female mimics in a lekking shorebird. *Biol Lett* 2006, **2**:161–164.
- Van Rhijn JG: Behavioural dimorphism in male ruffs *Philomachus pugnax* (L.). *Behaviour* 1973, **47**:153–229.
- Van Rhijn JG: *The Ruff*. London: Poyser; 1991.
- Lank DB, Smith CM, Hanotte O, Burke T, Cooke F: Genetic polymorphism for alternative mating behaviour in lekking male ruff, *Philomachus pugnax*. *Nature* 1995, **378**:59–62.
- Lank DB, Coupe M, Wynne-Edwards KE: Testosterone-induced male traits in female ruffs (*Philomachus pugnax*): autosomal inheritance and gender differentiation. *Proc R Soc Lond B* 1999, **266**:2323–2330.
- Lank DB, Farrell LL, Burke T, Piersma T, McRae SB: A dominant allele controls development into female mimic male and diminutive female ruffs. *Biol Lett* 2013. doi:10.1098/rsbl.2013.0653.
- Dale J, Lank DB, Reeve HK: Signalling individual identity versus quality: a model and case studies with ruffs, queleas, and house finches. *Am Nat* 2001, **158**:75–86.
- Lank DB, Dale J: Visual signals for individual identification: the silent "Song" of ruffs. *Auk* 2001, **118**:759–765.
- Hugie DM, Lank DB: The resident's dilemma: a female choice model for the evolution of alternative mating strategies in lekking male ruffs (*Philomachus pugnax*). *Behav Ecol* 1997, **2**:218–223.
- Widemo F: Alternative reproductive strategies in the ruff *Philomachus pugnax*: a mixed ESS? *Anim Behav* 1998, **56**:329–333.
- Lank DB, Smith CM, Hanotte O, Ohtonen A, Bailey S, Burke T: High frequency of polyandry in a lek mating system. *Behav Ecol* 2002, **2**:209–215.
- Farrell LL, Burke T, Slate J, Lank DB: A first-generation microsatellite linkage map of the ruff. *Ecol Evol* 2013. doi:10.1002/ece3.830.
- Green P, Falls K, Crooks S: *CRIMAP Documentation, version 2.4*. St. Louis: Washington University School of Medicine; 1990.

- Jaatinen K, Lehtikainen A, Lank DB: Female-biased sex ratios and the proportion of cryptic male morphs of migrant juvenile ruffs (*Philomachus pugnax*) in Finland. *Orn Fenn* 2010, **87**:125–134.
- Karlionova N, Pinchuk P, Meissner W, Verkuil Y: Biometrics of ruffs *Philomachus pugnax* migration in spring through southern Belarus with special emphasis on the occurrence of 'faeders'. *Ring Migr* 2007, **23**:134–140.
- Verkuil Y, Jukema J, Gill JA, Karlionova N, Melter J, Hooijmeijer JCEW, Peirsma T: Non-breeding faeder ruffs *Philomachus pugnax* associate according to sex, not morphology. *Bird Study* 2008, **55**:241–246.
- Dawson DA, Burke T, Hansson B, Pandhal J, Hale MC, Hinten GH, Slate J: A predicted microsatellite map of the passerine genome based on chicken-passerine sequence similarity. *Mol Ecol* 2006, **5**:1299–1320.
- Dawson DA, Åkesson M, Burke T, Pemberton JM, Slate J, Hansson B: Gene order and recombination rate in homologous chromosome regions of the chicken and a passerine bird. *Mol Biol Evol* 2007, **7**:1537–1552.
- Sinervo B, Bleay C, Adamopoulou C: Social causes of correlational selection and the resolution of a heritable throat color polymorphism in a lizard. *Evolution* 2001, **55**:2040–2052.
- Shuster SM, Sassaman C: Genetic interaction between male mating strategy and sex ratio in a marine isopod. *Nature* 1997, **388**:373–377.
- Farrell LL, Dawson DA, Horsburgh GJ, Burke T, Lank DB: Isolation, characterization and predicted genome locations of ruff (*Philomachus pugnax*, AVES) microsatellite loci. *Cons Gen Res* 2012, **4**:763–771. doi:10.1007/s12686-012-9639-0.
- Ducresc AL, Keller L, Roulin A: Pleiotropy in the melanocortin system, coloration and behavioral syndromes. *Trends Ecol Evol* 2008, **23**:502–510.
- Roulin A, Bize P, Ravussin P, Broch L: Genetic and environmental effects on the covariation between colour polymorphism and a life-history trait. *Evol Ecol Res* 2004, **6**:1253–1260.
- Silva AD, van den Brink V, Emaresi G, Luzio E, Bize P, Dreiss AN, Roulin A: Melanin-based colour polymorphism signals aggressive personality in nest and territory defence in the tawny owl (*Strix aluco*). *Behav Ecol Sociobiol* 2013, **67**:1041–1052.
- Anderson NG, Nagaeva O, Mandrika I, Petrovska R, Muceniece R, Mincheva-Nilsson LM, Wikberg JES: MC₁ receptors are constitutively expressed on leucocyte subpopulations with antigen presenting and cytotoxic functions. *Clin Exp Immunol* 2001, **3**:441–446.
- Hoekstra HE: Genetics, development and evolution of adaptive pigmentation in vertebrates. *Heredity* 2006, **97**:222–234.

doi:10.1186/1471-2156-14-109

Cite this article as: Farrell et al.: Genetic mapping of the female mimic morph locus in the ruff. *BMC Genetics* 2013 **14**:109.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

